Preliminary communication

Degradation during the hydrazinolysis of 2-acetamido-1-N-acyl-2-deoxy-β-D-glucopyranosylamines

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Hydrazinolysis is a convenient reaction for the isolation of the asparagine-bound carbohydrate moieties of glycoproteins¹. It also effects the *N*-deacetylation of *N*-acetylhexosamine residues, and the resulting glycan amine can be degraded regiospecifically by the nitrosation procedure². Excessively vigorous conditions can cause degradation during the hydrazinolysis and two sets of conditions are currently used, namely heating an anhydrous hydrazine solution in a sealed tube at 100° for 30 h (Bayard–Montreuil conditions³), and at 105° for 10 h in the presence of hydrazinium sulphate as a catalyst (Kochetkov conditions⁴).

Previous work⁵ has established that the major product of the hydrazinolysis of 2-acetamido-1-N-acetyl-2-deoxy- β -D-glucopyranosylamine, under Kochetkov conditions, was 2-amino-2-deoxy-D-glucose hydrazone. We now report the identification of a degradation product that is formed in this reaction.

As part of a study⁶ of the behaviour of saccharides under the conditions of hydrazinolysis, it was found that 2-amino-2-deoxy-D-glucose hydrochloride was converted, via 2-amino-2-deoxy-D-glucose hydrazone, into 1-deoxy-D-fructose hydrazone under Bayard—Montreuil conditions. The 1-deoxy-D-fructose hydrazone was identified by ^{1}H (Me singlet at δ 1.84) and ^{13}C (Me at 13.8 p.p.m.) n.m.r. measurements, by comparison with the hydrazone prepared from authentic 1-deoxy-D-fructose⁷, and by sequential nitrosation and reduction with sodium borodeuteride, to give a mixture of 1-deoxy-D-[2- ^{2}H]glucitol and -D-[2- ^{2}H] mannitol which were identified by g.l.c.-m.s. measurements on the peracetates (m/z 88, MeCDOAc 4).

The product of hydrazinolysis of 2-acetamido-1-N-acetyl-2-deoxy-β-D-glucopyranosylamine under Kochetkov conditions contained 22% of 1-deoxy-D-fructose hydrazone, estimated from the p.m.r. spectrum. However, under Bayard—Montreuil conditions, amide cleavage was very inefficient; starting material (35%) and 1-N-acetyl-2-amino-2-deoxy-β-D-glucopyranosylamine (35%) were detected in the product by n.m.r. spectroscopy. This result was surprising in view of the recent report that a reaction time of only 8–12 h is sufficient to effect the complete release of the asparagine-bound oligosaccharides in glycoproteins and glycopeptides. We considered that the hydrazinolysis of glycoproteins and

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Scheme 1

glycopeptides might involve a neighbouring-group reaction, and this was confirmed by the hydrazinolysis of 2-acetamido-1-*N*-(L- β -aspartyl)-2-deoxy- β -D-glucopyranosylamine (GlcNAc-Asn). Under Kochetkov conditions, 1-deoxy-D-fructose hydrazone was formed in ~44% yield; under Bayard Montreuil conditions, amide cleavage was complete and the yield of 1-deoxy-D-fructose hydrazone was lower (~14%), presumably because its precursor, 2-amino-2-deoxy-D-glucose hydrazone, was released more slowly than under Kochetkov conditions. The more efficient amide-cleavage in GlcNAc-Asn than in 2-acetamido-1-*N*-acetyl- β -D-glucopyranosylamine must be due to an intramolecular nucleophilic catalysis, as depicted in Scheme 1.

In the hydrazinolysis of glycoproteins and glycopeptides in which asparagine is linked through its amino group to another amino acid, the hydrazide 1 is a possible intermediate in the reaction. Participation by the hydrazide nitrogen atom as shown (1) is then possible.

The formation of 1-deoxy-D-fructose hydrazone from 2-amino-2-deoxy-D-glucose hydrazone can be envisaged as proceeding via the 1,2-bishydrazone, whose formation is analogous to the formation of phenylosazone derivatives. A Wolff-- Kishner type reaction at

C-1, possibly facilitated by an intramolecular process involving a six- or seven-membered ring transition-state as depicted in Scheme 2, would then give 1-deoxy-D-fructose hydrazone. A similar reaction at C-2 would give 2-deoxy-D-arabino-hexose hydrazone, and it is interesting to note that a 2-deoxy-D-arabino-hexitol derivative was recently identified⁸ as a minor product of the sequential hydrazinolysis, nitrosation and reduction of glycopeptides.

Our studies of these reactions continue, and the significance of the results in the context of structural studies of glycoproteins will be discussed elsewhere.

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